FINAL DRAFT REPORT

Intermedia Transfer Factors for Contaminants Found at Hazardous Waste Sites

TRICHLOROETHYLENE (TCE)

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Prepared for:

The Office of Scientific Affairs

The Department of Toxic Substances Control (DTSC)

and the California Environmental Protection Agency
in Support of the CalTOX Model

December 1994

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FORWARD

The Department of Toxic Substances Control (DTSC), within the California Environmental Protection Agency, has the responsibility for managing the State's hazardous-waste program to protect public health and the environment. The Office of Scientific Affairs (OSA) within the DTSC provides scientific assistance in the areas of toxicology, risk, environmental assessment, training, and guidance to the regional offices within DTSC. Part of this assistance and guidance is the preparation of regulations, scientific standards, guidance documents, and recommended procedures for use by regional staff, local governmental agencies, or responsible parties and their contractors in the characterization and mitigation of hazardous-waste-substances-release sites. The CalTOX model has been developed as a spreadsheet model to assist in exposure and health-risk assessments that address contaminated soils and the contamination of adjacent air, surface water, sediments, and ground water.

The modeling effort includes multimedia transport and transformation models, exposure scenario models, and efforts to quantify and reduce uncertainty in multimedia, multiple-pathway exposure models. Use of the CalTOX model requires that we determine the intermedia transfer factors (ITFs) that define concentration relationships between an exposure medium and the environmental medium that is the source of the contaminant. ITFs are chemical and physical parameters which serve as inputs in the CalTOX model analysis.

This report provides a set of ITFs needed to run the CalTOX model for TCE. For this chemical, we have conducted a critical review of existing literature for measured values and estimation methods in order to compute an arithmetic mean (\bar{x}) , a coefficient of variation (CV), and plausible range for each ITF.

OVERVIEW

The purpose of this report is to provide a set of chemical-specific intermedia-transfer factors (ITFs) for TCE. We have carried out a critical review of the existing literature in order to identify a mean value, coefficient of variation (CV) and value range for the ITFs listed in Table 1. For values used to define a given parameter, our highest priority was given to experimental values reported in the primary scientific literature, that is, peer-reviewed journals. For parameters that are not readily available from the primary literature, widely cited secondary references such as Lyman et al. (1982, 1990), Verschueren (1984), Howard et al. (1990, 1991), Mackay et al. (1992), the CRC Handbook (1989-90) and the Merck Index (1983, 1989) are used to establish parameter values. When measured values are not available from either the primary literature or secondary references, estimates of ITF parameter values are based on estimation equations that are available in the primary literature. Typically, these estimation methods relate ITFs to other measured contaminant parameters using quantitativestructure-activity-relationship (QSAR) methods. In these cases, parameter values estimated from a QSAR method are treated as the arithmetic mean and the estimation error of the method is used to determine the CV. Table 1 summarizes the units required by the CalTOX model, the values of chemical specific physico-chemical properties, distribution coefficients, biotransfer and bioconcentration factors, and transformation half-lives obtained in this study.

CalTOX Chemical-Specific Input Requirements

The CalTOX model uses three sets of input data—one describing the chemical-specific properties of the contaminants, a second providing properties of the environment or landscape receiving the contaminants, and a third that defines for exposure assessment the characteristics of individuals in various age/sex categories and the characteristics of the micro-environments in which they live or from which they obtain water and food. Each of the inputs in these sets must be described in terms of a mean value with an estimated coefficient of variation, which describes the uncertainty or variability associated with that parameter. This report addresses mean value, CV, and range of values needed to characterize chemical-specific inputs.

Physicochemical Properties

Physicochemical properties include molecular weight, octanol-water partition coefficient, melting point, vapor pressure, Henry's law constant, diffusion coefficients in air and water, and the organic-carbon partition coefficient. The octanol-water partition coefficient provides a measure of the extent of chemical partitioning between water and octanol at equilibrium and is used as a basis for estimating other ITF parameters. The melting point is the temperature at which a compound makes the transition from a solid to a liquid phase. Vapor pressure is the pressure exerted by a chemical vapor in equilibrium with its solid or liquid phase. Water solubility is the upper limit on a chemical's dissolved concentration in pure water, at a specified temperature.

Table 1. Summary of Chemical Properties for Trichloroethylene

Description	Symbola	Mean Value	Coefficient of Variation	Number of Values
Molecular Weight (g/mol)	MW	131.4	3.9×10^{-4}	5
Octanol-Water Partition Coefficient	K _{ow}	320	0.32	6
Melting Point (K)	T _m	189.7	0.026	7
Vapor Pressure (Pa)	VP	9700	0.021	5
Solubility (mol/m ³)	S	11	0.15	7
Henry's Law Constant (Pa-m ³ /mol)	Н -	890	0.18	12
Diffusion Coefficient in Pure Air (m ² /d)	Dair	0.68	0.05	e
Diffusion Coefficient in Pure Water (m ² /d)	D _{water}	9.0×10^{-5}	0.25	e
Organic Carbon Partition Coefficient	K _{oc} -	86	0.46	13
Distribution Coefficient in Ground-Surface and Root-Zone Soil	K _{d_s} -	b	e	e
Distribution Coefficient in Vadose-Zone Soil	K _{d_v} -	b	e	e
Distribution Coefficient in the Ground-Water Zone	K _{d_q} -	b	e	e
Distribution Coefficient in Ground Water Sediment	K _{d_d} -	b	e	e
Partition Coefficient in Plants Relative to Soil Concentration [ppm (pFM) /ppm (sFM)]	K _{ps} -	0.25	4.0	e
Biotransfer Factor in Plants Relative to Contaminant Air Concentration (m ³ [a]/kg[pFM])	K _{pa} -	0.011	14	e
Biotransfer Factor in Milk Relative to Cattle-Diet Contaminant Intake (d/kg)	B _k -	2.8 × 10 ⁻⁶	11	e
Biotransfer Factor in Meat Relative to Cattle-Diet Contaminant Intake (d/kg)	B _t -	2.5×10^{-5}	13	e
Biotransfer Factor in Eggs Relative to Hen-Diet Contaminant Intake (d/kg)	B _e -	0.0026	14	e
Biotransfer in Breast Milk Relative to Contaminant Intake by the Mother (d/kg)	B _{bmk} -	$\boxed{6.4\times10^{-5}}$	10	e
Bioconcentration Factor in Fish Relative to Contaminant Water Concentration	BCF -	53	1.0	2
Skin Permeability Coefficient (cm/h)	К _{р_w} -	0.047	2.4	e
Skin-Water/Soil Partition Coefficient [ppm (skin)/ppm (water)]	K _m -	26	0.27	e
Reaction Half-Life in Air (d)	T _{half_a}	3.5	0.11	6
Reaction Half-Life in Ground-Surface Soil (d)	T _{half_g}	930	1.7	3
Reaction Half-Life in Root-Zone Soil (d)	T _{half_s}	930	1.7	3
Reaction Half-Life in the Vadose-Zone Soil (d)	T_{half_v}	760	1.4	6
Reaction Half-Life in Ground-Water Zone Soil (d)	T_{half_q}	800	1.5	5
Reaction Half-Life in Surface Water (d)	T_{half_w}	120	0.88	7
Reaction Half-Life in the Sediment (d)	T _{half_d}	220	0.67	4
a Values followed by a "-" include default equations that can be used for estimations b K $_d$ = [(K $_0$ c) × (fraction organic matter)], a site and soil zone specific parameter e estimated parameter value				

Henry's law constant is a measure at equilibrium of the ratio of chemical activity in the gas above a liquid to chemical activity in the liquid. Diffusion coefficients describe the movement of a molecule in a liquid or gas medium as a result of differences in concentration within the medium. They are used to calculate the dispersive component of chemical transport. The higher the diffusion coefficient, the more likely a chemical is to move in response to concentration gradients. The organic-carbon partition coefficient provides a measure of chemical partitioning between organic carbon (in soils, rocks, and sediments) and water. The higher the $K_{\rm oc}$, the more likely a chemical is to bind to the solid phase of soil or sediment than to the liquid phase.

The Solid-Water Distribution Coefficients

The distribution or sorption coefficient, K_d , is the concentration ratio, at equilibrium, of chemical attached to solids and/or particles (mol/kg) to chemical concentration in the solution, mol/L. When K_{oc} is multiplied by the fraction organic carbon in a soil or sediment, we obtain an estimate of the soil/water or sediment/water partition coefficient. CalTOX requires, as input, distribution coefficients for ground-surface, rootzone, and vadose-zone soil; ground-water-zone rock or soil, and surface-water sediments.

Biotransfer Factors and Bioconcentration Factors

The CalTOX model requires, as input, general relationships that can be used to estimate partition coefficients between air and plants; between soil and plants; between animal feed intake and animal-based food products; between surface water and fish; between the human mother's uptake and breast milk; between skin and water; and between skin uptake and concentration in skin water.

The chemical-specific plant-air partition coefficient, K_{pa} , represents the ratio of contaminant concentration in above-ground plant parts, in mg/kg (fresh mass), to contaminant concentration in the gas-phase of the atmosphere mg/m³ (air). The plant-soil partition coefficient, K_{ps} , expresses the ratio of contaminant concentration in plant parts, both pasture and food, in mg/kg (plant fresh mass) to concentration in wet root-zone soil, in mg/kg.

The biotransfer factors B_t , B_k and B_e are the steady-state contaminant concentrations in, respectively, fresh meat, milk, and eggs; divided by the animals' daily contaminant intake. These factors are expressed in units of (mg/kg)/(mg/d), or kg/d. Unlike bioconcentration factors, which express steady-state concentration ratios between animal tissue and a specific environmental medium, biotransfer factors express the steady-state relationship between intake and tissue or food-product concentrations.

Lactating women can transfer to breast milk their intake of contaminants from all intake routes—ingestion, inhalation, and dermal contact. B_{bmk} is the biotransfer factor for milk-concentration versus the mother's intake. This relationship may also be

described as the ratio of contaminant concentration in mother's milk divided by the mother's daily intake of that contaminant, in units of d/kg (milk).

The bioconcentration factor BCF provides a measure of chemical partitioning between fish tissue based on chemical concentration in water.

Chemical specific exposure factors used in CalTOX include the skin-water and skin-soil partition coefficients. K_m is the skin-water partition coefficient in cm³ (water)/cm³ (skin). In order to estimate the skin-soil partition factor, K_m^{soil} , with units cm³(soil)/cm³(skin), we divide equation K_m by the sorption coefficient K_d for soil, or

$$K_{\rm m}^{\rm soil} = \frac{K_{\rm m}}{K_{\rm d}}$$

 K_{p_w} is the steady-state permeability coefficient in cm/hour for a contaminant from water on skin through stratum corneum and can either be based on a measured value or estimated values.

Chemical-Specific Transformation Process Half-Lives

Chemical transformations, which may occur as a result of biotic or abiotic processes, can have a profound effect on the persistence of contaminants in the environment. Experimental methods and estimation methods are available for defining these fate processes in a variety of media. Specific information on the rates and pathways of transformation for individual chemicals of concern should be obtained directly from experimental determinations, if possible, or derived indirectly from information on chemicals that are structurally similar. CalTOX makes use of media- and reaction-specific reaction half-lives to establish rate constants for transformation removal processes that include photolysis, hydrolysis, oxidation/reduction, and microbial degradation.

Transformation-rate half-lives are among the more uncertain parameters in the CalTOX model. There are typically few available measurements or ranges of estimated values in the primary and secondary literature. Most of the available half-life values are obtained from limited measurements for environmental media that are not necessarily representative of those in California. These values often involve scientific judgment as much as measurement. In making use of these data, we expanded the range of the reported values by a factor of 5 when only 2 or 3 representative values are presented and by a factor of 10 when only one value is provided. If 4 or more measured values are available, these uncertainty factors are not applied. In order to express the lack of reliability associated with a limited number of measured values for a parameter, these uncertainty factors are used to express both large uncertainty and significant variability.

Statistical Methods

Each of the inputs to CalTOX must be described by a mean value and an estimated coefficient of variation which describes the uncertainty or variability associated with that parameter. For input values that are derived from a number of measured values, the mean and coefficient of variation are obtained from the arithmetic mean and the arithmetic standard deviation of the inputs. For estimated input values, the mean and coefficient of variation are obtained from an estimation equation and the residual error of the estimation equation. The methods we used to obtain these values are described here.

Mean and Coefficient of Variation

The arithmetic mean (\bar{x}) is used to represent all inputs that are derived from a number of measured values—even those that might have geometric distributions. The (\bar{x}) is computed by summing the reported values and dividing this sum by the total number of observations:

$$\begin{array}{c} n \\ X_i \\ Arithmetic\ mean\ (\overline{x}) = \frac{i=1}{n} \end{array}$$
 (Eqn. 1)

Where x_i is the sum of the observed values and n is the number of observations. In this case, the coefficient of variation (CV) is computed by dividing the arithmetic standard deviation (x_n) by the mean. Standard deviation and CV are computed according to the following equations:

standard deviation
$$(\mathbf{S_n}) = \sqrt{\frac{\frac{n}{(\mathbf{x_i} - \bar{\mathbf{x}})^2}}{n}}$$
 (Eqn. 2)

coefficient of variation (CV) =
$$\frac{s_n}{\bar{x}}$$
 (Eqn. 3)

It should be noted that, based on the central limit theorem of statistics, the confidence associated with the estimate of $\bar{\mathbf{x}}$ from above becomes large as the number of samples used to estimate $\bar{\mathbf{x}}$ also becomes large. Therefore, the reliability of the estimates of mean and CV of a parameter are low when the sample size is small. It is beyond the scope of this document to explicitly address the reliability of these estimates. Nonetheless, in order to give an indication of potential reliability problems, we list the number of measurements used to estimate the mean and CV of each parameter in the last column of Table 1.

Estimation Equations and the Residual Errors of the Estimation Method

Estimates of some CalTOX inputs are based on regression equations that relate a parameter value to some measure of structure or activity associated with the contaminant. These methods are referred to as quantitative structure-activity relationship (QSAR) methods. The reliability of a parameter-value estimated in this way is defined by the precision of these QSAR methods.

Our estimate of precision in QSAR estimation methods is based on calculating, S_e , the standard error of the estimate (or standard deviation of the residuals). This error calculation is based on the regression equations and fragment models used to derive a parameter value. To illustrate, when the value of parameter such as the organic-carbon partition coefficient (K_{oc}) is estimated using a regression or correlation analysis, the S_e is calculated using the following approach (Hamburg, 1970). First, since it is typical that it is the log K_{oc} (not K_{oc} itself) that is estimated from a regression equation, we calculate the S_e of log K_{oc} according to

$$S_{e} \text{ of log } K_{oc}^{est} = \sqrt{\frac{\left(\log K_{oc}^{msd} - \log K_{oc}^{est}\right)^{2}}{(n-2)}}$$
(Eqn. 4)

where n is the number of chemicals used in the estimation protocol and K_{oc}^{est} refers to the estimated property (K_{oc} in this case) and K_{oc}^{msd} refers to the corresponding measured values used to carry out the regression. In order to calculate the S_e of K_{oc} we make use of the transformation

GSD
$$(K_{oc}^{est}) = 10^{(Se \text{ of } log K_{oc}^{est})}$$
 (Eqn. 5)

to calculate the geometric standard deviation of S_e (GSD) of K_{oc}^{est} , which is simply the GSD of the K_{oc} estimate, that is GSD (K_{oc}^{est}). It has been shown by Atchison and Brown (1957) that the relationships between the GSD and CV for log normal distributions are as follows

$$GSD = \exp\left\{\sqrt{\ln(1+CV^2)}\right\}$$
 (Eqn. 6)

$$CV = \sqrt{\left(\exp\left\{ \left[\ln(GSD)\right]^2\right\} - 1\right)}$$
 (Eqn. 7)

Since the implicit assumption of a regression for estimating the log of K_{oc} is that any estimated value, log (K_{oc}^{est}) , is centered on normal distribution with standard deviation equal to S_e of log K_{oc} it follows that the corresponding estimated value of K_{oc} is centered on a log normal distribution with GSD (K_{oc}^{est}) and with

$$CV (K_{oc}^{est}) = \sqrt{exp \left\{ \left[ln(GSD(K_{oc}^{est})) \right]^2 \right\} - 1}$$
 (Eqn. 8)

This approach is used to estimate CVs for the estimation equations presented in this document.

In some cases the error term, CV for example, is calculated by combining through the operations of multiplication and division the CVs of two or more parameters. For example the CV in the ration H = VP/S is combined from the CV (VP) and CV (S). In this case, if the input parameters are independent, the combined CV is calculated using the following equation:

$$CV_{\text{combined}} = \sqrt{\frac{\sum_{i=1}^{n} CV_{i}^{2}}{n}}$$
 (Eqn. 9)

where n is the number of parameters used in the multiplication/division and CV_i is the coefficient of variation in the *i*th input parameter.

Trichloroethylene (TCE)

Other Names

acetylene trichloride, 1-chloro-2,2-dichloroethylene, 1,1-dichloro-2-chloroethylene, 1,1,2-trichloroethylene, ethinyl trichloride, ethylene trichloride, TCE, Algylen, Benzinol, Blacosolv, Cecolene, Chlorilen, Chlorylen, Circosolv, Densinfluat, Dow-Tri, Dukeron, Fleck-Flip, Flock-Flip, Fluate, Gemalgene, Germalgene, Lanadin, Lethurin, Narcogene, Narkosoid, Nialk, Perm-A-Clor, Petzinol, Philex, Threthylen, Trethylene, Trial, TRI, Triad, Triasol, Trichloran, Trichloren, Trichloroethene, Triclene, Tri-Clene, Trielene, Trielin, Trielina, Triklone, Trilen, Trilene, Triline, Trimar, Triol, Tri-Plus, Tri-Plus M, Vestrol, Vitran, Westrosol (WHO IARC, 1979).

Background

Trichloroethylene (TCE) is not known to occur as a natural product. It is commercially produced by chlorination and dehydrochlorination of 1,2-dichloroethane and belongs to the chemical family of chlorinated alkenes. Major use of TCE is in vapor degreasing of fabricated metal parts. It is often used as a carrier solvent in textile cleaning and solvent extraction processes, as a lubricant and adhesive and as a low-temperature heat transfer fluid. TCE is also used in the production of polyvinyl chloride (PVC), paints, coatings and some miscellaneous chemical synthesis. It is estimated that 60-90% of the world TCE production is released into the environment and volatilization appears to be the primary transport process for removal. Once in the troposphere, it reacts with hydroxyl radical. TCE contamination in the aquatic environment will result in some bioaccumulation in marine organisms (IPCS, 1984).

Formula

C₂ H Cl₃

$$CI$$
 $C = C$ CI CI

MW: Molecular Weight

The units used for molecular weight are grams/mole (g/mol).

Reported Values

131.38	reported by Sax and Lewis (1989)
131.388	reported by Daubert and Danner (1989)
131.389	reported by Holden (1980) [also cited in Riddick et al. (1986)]
131.39	reported in CRC Handbook [Weast et al. (1989)]
131.5	reported by Verscheuren (1983)

From the above 5 reported values above, we obtain the following statistics for the molecular weight of TCE:

Arithmetic mean (coefficient of variation): $MW = 131.4 \; (3.9 \times 10^{-4}) \; g/mol$

Kow: Octanol-Water Partition Coefficient

The units used for K_{ow} are $\frac{mg/liter\ (octanol)}{mg/liter\ (water)}$ and K_{ow} is therefore unitless.

Experimental Values

195	reported as a log $K_{ow} of 2.29$ by Glave and Hansch [unpublished results in Hansch and Leo (1987)]
263	reported at 20 $^{\circ}\text{C}$ as a log K_{ow} of 2.42 by Veith et al. (1980) estimated using reverse-phase liquid chromatography
407	reported at 20°C as a log $K_{\rm ow} {\rm of} 2.61$ reported by Chiou and Freed (1977) using shake flask-gas chromatography
263	reported at 23 ± 1.5 °C as a K_{ow} of 263 by Banerjee et al. (1980) using a shake-flask LSC method [also cited in Mackay et al. (1993)]
468	reported at 23 °C as a log $K_{\rm ow}$ of 2.67 reported by Harnisch et al. (1983) estimated using the HPLC method (extrapolated to 100% water phase)
339	reported at 25 $^{\circ}$ C as a log K $_{\rm ow}$ of 2.53 by Tewari et al. (1982) estimated using a generator column-HPLC method [also cited by Mackay et al. (1993)]

From the 6 measured values above we obtain the following statistics for the octanol-water partition coefficient of TCE at 20 to 25 °C:

Arithmetic mean (coefficient of variation): $K_{ow} = 320 (0.32)$

Range: $K_{ow} = 195 \text{ to } 468$

T_m: Melting Point

The units used for melting point are kelvins (K).

Reported Values

186	reported as a MP of -87 °C by Verscheuren (1983)
186	reported as a MP of -86.8 °C by Clayton and Clayton [Patty's Industrial Hygiene and Toxicology (1981)]
187	reported as a MP of -86.4 °C by McGovern (1943)
188	reported as a MP of -84.8 °C by Budavari et al. [Merck Index (1989)]
188.4	reported as a MP of 188.4 K by Daubert and Danner (1989)
191	reported as a MP of -81.7 °C by Kirk-Othmer (1985)
200	reported as a MP of -73 °C by Weast et al. [CRC Handbook (1989)]

From the 7 measured values above, we obtain the following statistics for the melting point of TCE:

Arithmetic mean (coefficient of variation): $T_m = 189.7 (0.026) \text{ K}$

Range: 186 to 200 K

VP: Vapor Pressure at Standard Temperatures

The units used for vapor pressure are pascals (Pa).

Experimental Values

9466	reported at 25 °C as 71 mm Hg by McGovern (1943)
9546	reported at 25.5 $^{\circ}\text{C}$ as 71.6 mm Hg by McDonald (1944) [also cited by Boublik et al. (1984)]
9599	reported at 25 °C as 72 mm Hg by Sconce et al. (1962)

9733 reported at 25 °C as 9.733 kPa by Hertz and Rathmann (1912)

[also cited in Boublik (1984)]

extrapolated to 25 °C as 74.9 mm Hg by Stull (1947) using an Antoine

equation and -22.8 < T < 48 °C

From the 5 measured values above, we obtain the following statistics for the vapor pressure of TCE at 25 °C:

Arithmetic mean (coefficient of variation):

$$VP = 9700 (0.021) Pa$$

Range: 9466 to 9986 Pa

Estimation Methods

The following Antoine equations have been published for estimating the vapor pressure of TCE in kPa

Antoine Equation 1

The following Antoine equation is derived from the above data by Stull (1947).

$$log_{10} VP (Pa) = 10.0944 - \frac{1816.26}{273 + T (^{\circ}C)}$$

$$VP = 9990 Pa at 298 K$$

Antoine Equation 2

The following Antoine equation was derived from data by Kirk-Othmer (1964).

$$log_{10} VP (Pa) = 10.128 - \frac{1830.4}{273 + T (^{\circ}C)}$$

for -20
$$^{\circ}\text{C} < T < 60$$
 $^{\circ}\text{C}, \, yielding$

S: Solubility in Water

The units used in the solubility values below are $\frac{mg}{liter (water)}$ (mg/L).

Experimental Values

1100	reported at 25 °C as 0.11 g/100 g water by McGovern (1947)
1367	reported at 25 $^{\circ}$ C as 0.0104 mol/L by Tewari et al. (1982) estimated using an HPLC method [also cited by Riddick et al. (1986)]
1421	reported at 23-24°C by Broholm et al. (1992) using shake bottle-GC analysis
1472	reported at 25 °C as 11200 μM by Banerjee et al. (1980) using a shake-flask LSC method [also cited in Horvath (1982)]
1474	reported at 25 °C as a log 1/solubility of 1.95 mol/L by Veith et al. (1980) using a stir flask method [also cited by Mackay et al. (1993)]
1500	reported at 20 $^{\circ}\text{C}$ as 0.150 g/100 ml reported by Chiou and Freed (1977) using a shake-flask GC method
1818	reported at 25 °C as 1 g/550 ml from Wright and Schaffer (1932) in Horvath (1982) [also cited by Mackay et al. (1993)]

Unit Conversion

Arithmetic mean (coefficient of variation) of TCE solubility

= 1450 (0.15) mg/L

 $= 11 (0.15) \text{ mol/m}^3$

From the 7 measured values above, we obtain the following statistics for the water solubility of TCE at 25 $^{\circ}\text{C}:$

Arithmetic mean (coefficient of variation):

 $S = 11 (0.15) \text{ mol/m}^3$

Range: 8 to 11 mol/m³

H: Henry's Law Constant

The units used for Henry's Law constant are $\frac{Pascals-m^3}{mole}$ (Pa-m³/mol).

Experimental Values

reported at 20 °C as 0.00674 atm-m³/mol by Lincolff and Gossett (1984) using a batch air stripping method

709	reported at 20 $^{\circ}$ C as 0.0070 atm-m ³ /mol by Tse (1992) estimated using a GC method
774	reported at 20 °C as 0.00764 m³-atm/mol by Lincolff and Gossett (1984) using Equilibrium Partitioning in a Closed System (EPICS)
805	reported at 20 °C as 0.325 unitless by Lee (1990) using EPICS
830	reported at 20 $^{\circ}$ C as 0.335 dimensionless by Munz & Roberts (1987) using the multiple equilibration method [also cited in Mackay et al. (1993)]
853	reported at 20 °C as 0.00842 atm-m 3 /mol by Ashworth (1988) using EPICS
1065	reported at 20 °C as 0.43 (unitless) by Yurteri et al. (1987) using EPICS
768	reported at 25 °C as 0.31 (unitless) by Tancréde and Yanagisawa (1990) using purge and trap gas chromatography [also cited in Mackay et al. (1993)]
962	reported at 25 °C as 0.397 (unitless) by Garbarini and Lion (1985) using an equilibrium headspace technique (EPICS)
971	reported at 25 °C as 0.00958 atm-m³/mol by Gossett (1987) using EPICS [also cited in Mackay et al. (1993)]
1034	reported at 25 °C as 0.0102 atm-m³/mol by Ashworth (1988) using EPICS
1186	reported at 25 °C as 0.0117 atm-m 3 /mole by Warner et al. (1987) using a modified batch-stripping apparatus

From the 12 measured values above, we obtain the following statistics for the Henry's law constant in the range of 20-25 °C:

Arithmetic mean (coefficient of variation):

$$H = 890 (0.18) Pa-m^3/mol$$

Range: 683 to 1186 Pa-m³/mol

Estimation Method

$$H = {VP (Pa) \over S (mol/m^3)} = {9666 \over 11} = 880 (0.10) Pa-m^3/mol$$

Dair: Diffusion Coefficient in Pure Air

The units used for the diffusion coefficient in pure air are $\frac{meters^2}{day}$ (m²/d).

Estimation Method

Based on the Fuller et al. (1966) method described in Lyman et al. (1982), the estimated diffusion coefficient in air (m^2/d) is given by:

$$D_{air} = 8.6 \times 10^{-3} \text{ T}^{1.75} \frac{\sqrt{(29 + M_x)/(29 \times M_x)}}{\left[2.7 + V_x^{1/3}\right]^2}$$

Molar volume (V_x) can be estimated by the LeBas incremental method as described in Lyman et al. (1982) With a molar volume, V_x , of 107 cm³/mol, molecular weight (M_x) of 131 g/mol, and a temperature equal to 298 K, the above expression gives:

$$D_{air} = 3.2 \times 10^{-5} \ T^{1.75} = 0.68 \ m^2/d$$

The reported average absolute estimation error is 5 to 10% (Fuller et al., 1966). This estimation error is estimation error is <5% in chlorinated aliphatics and is equivalent to the CV reported below.

Based on the estimated value and the estimation error reported above, we obtain the following statistics for the estimated air diffusion coefficient of TCE at 25 $^{\circ}$ C:

Arithmetic mean (coefficient of variation): $D_{air} = 0.68 (0.05) \text{ m}^2/\text{d}$

Dwater: Diffusion Coefficient in Pure Water

The units used for the diffusion coefficient in pure water are $\frac{meters^2}{day}$ (m²/d).

Estimation Method

Based on the Wilke and Chang (1955) method described in Reid et al. (1987) the diffusion coefficient in water (m^2/d) is given by:

$$D_{water} = \frac{6.5 \times 10^{-7} \sqrt{f \times M_y} T}{h_y V_x^{0.6}}$$

Wilke and Chang (1955) recommend an association factor, f, of 2.6 when the solvent is water. The viscosity of water, h_y , is 0.89 cP at 25 °C. Molar volume (V_x) can be estimated by the LeBas incremental method as described in Lyman et al. (1982). With a V_x equal to 107 cm³/mol, a temperature (T) of 298 K, and My (MW of water) equal to 18 g/mol., this expression gives:

$$D_{water} = 3.0 \times 10^{-7} T = 9.0 \times 10^{-5} m^2/d \text{ at } 25 \text{ }^{\circ}\text{C}$$

Original data for this estimation, provided in Reid et al. (1987), can be used to determine the standard error of the estimator for this estimation method. From this data we calculate a CV of 0.25 from a 25% estimation error.

Based on the estimated value and the estimation error reported above, we obtain the following statistics for the estimated water diffusion coefficient of TCE at 25 °C:

Arithmetic mean (coefficient of variation):

$$D_{water} = 9.0 \times 10^{-5} (0.25) \text{ m}^2/\text{d}$$

Koc: Organic-Carbon Partition Coefficient

The units used for K_{oc} are $\frac{mg/kg \text{ (organic carbon)}}{mg/kg \text{ (water)}}$ and K_{oc} is therefore unitless.

Experimental Values

- 18.5 reported at 25 °C as a $\rm K_{oc}$ of 14 to 23 by Liljestrand and Charbeneau (1987) using an organic clay soil from an aquifer (19 ft.) with a fraction organic carbon ($\rm f_{oc}$) of 0.169
- reported as an average K_d of 0.88 by Lee et al. (1989) using an untreated Martlette (A horizon) soil (pH = 6.4, f_{oc} = 2.59%)
- reported as a K_{om} of 33.1 by Rutherford & Chiou (1992) using a peat soil (f_{oc} = 57%) and assuming K_{oc} = $K_{om} \times 1.724$ [also cited in Mackay (1993)]
- reported as a soil/water K_p of 0.16 by Piwoni and Banerjee (1989) using soil (f_{oc} = 0.0026) for 23 hrs. at 10 to 300 μ g/L
- reported as an average log K_{oc} of 1.81 by Abdul et al. (1987) using a batch absorption method and sandy vadose-zone materials (f_{oc} = 1%)

73	reported as an average K_{oc} of 72.5 by Seip et al. (1986) using 3 surface soils (pH = 4.2-7.4, f_{oc} = 0.2-3.7%) and a concentration of 0.5 mg/l
87	reported as a $\rm K_d$ of 1.57 by Rogers and McFarlane (1981) from an Overton Silty Clay Loam in Southeastern Nevada ($\rm f_{oc}$ = 1.8%, pH = 7.8) and 100 ppb TCE
99	reported at 25 °C and equivalent to an average K_{oc} of 99 by Urano and Murata (1985) using 5 soils (f_{oc} = 0.1 to 4.9%) and a batch equilibration method [also cited in Mackay et al. (1993)]
101	reported as an average K_{om} of 58.3 by Friesel et al. (1984) and assuming that K_{oc} = $K_{om} \times 1.724$
106	reported as a K_{oc} of 106 measured by Garbarini & Lion (1986) using Sapsucker Woods Soil (f_{oc} = 7.51%) [also cited in Mackay et al. (1993)]
123	reported as a log $\rm K_{oc}$ of 2.09 in Grathwohl (1990) in soil, sand and loess and corrected to 20 $^{\circ}{\rm C}$ [also reported in Mackay (1993)]
140	reported as an average $\rm K_d$ of 2.7 by Doust and Huang (1992) using average data from 2 soils (average $\rm f_{oc}$ = 1.93%)
150	reported as a $\rm K_d$ of 3.89 by Rogers and McFarlane (1981) from a Hastings silty clay loam in Nebraska ($\rm f_{oc}$ = 2.6%, pH = 5.6) and 10 ppb TCE

From the 13 measured values above, we obtain the following statistics for the organic carbon partition coefficient for TCE:

Arithmetic mean (coefficient of variation):
$$K_{oc} = 86 (0.46)$$

Range: 18.5 to 150

Estimation Method

Karickhoff (1981) has described empirical estimation methods for obtaining K_{oc} from K_{ow} . The most general of these is that K_{oc} is equal to 0.41 times K_{ow} .

$$K_{oc} = 0.41 \times K_{ow}$$
$$K_{ow} = 320$$

$$K_{oc}$$
 (est) = 131 (1)

The reported CV is based on data provided by Karickhoff (1981). This estimation error does not include uncertainty in the value of K_{ow} .

Kd s: Distribution Coefficient in Ground-Surface and Root-Zone Soil

The units used for K_{d_s} are $\frac{mg/kg \text{ (dry surface and root-zone soil)}}{mg/kg \text{ (water)}}$ and K_{d_s} is therefore unitless.

Estimation Method

This is a site specific parameter and depends on the fraction organic carbon in the surface and root-zone soil and on the value of K_{oc} . K_{d_s} is the product of the soil organic carbon partition coefficient (K_{oc}) and the fraction organic carbon in the surface and root-zone soil (f_{oc_s}) (Karickhoff, 1981).

$$\begin{split} K_{d_s} &= K_{oc} \times f_{oc_s} \\ f_{oc_s} &= \frac{kg \ organic \ carbon \ (dry \ surface \ and \ root-zone \ soil)}{kg \ (soil)} \end{split}$$

Based on the estimation reported above, we obtain the following equation for the distribution coefficient in surface and root-zone soil. K_{d_s} is a site and soilzone specific parameter depending on the fraction organic carbon in the surface and root-zone soil or:

$$K_{d_s} = K_{oc} \times f_{oc_s}$$

K_{d_v}: Distribution Coefficient in Vadose-Zone Soil

The units used for K_{d_v} are $\frac{mg/kg \text{ (dry vadose-zone soil)}}{mg/kg \text{ (water)}}$ and K_{d_v} is therefore unitless.

Estimation Method

This is a site specific parameter and depends on the fraction organic carbon in the vadose-zone soil and on the value of K_{oc} . K_{d_v} is the product of the soil organic carbon partition coefficient (K_{oc}) and the fraction organic carbon in the vadose-zone soil (f_{oc} $_{v}$) (Karickhoff, 1981).

$$K_{d_v} = K_{oc} \times f_{oc_v}$$

$$f_{oc_v} = \frac{kg \ organic \ carbon \ (dry \ vadose-zone \ soil)}{kg \ (soil)}$$

Based on the estimation reported above, we obtain the following equation for the distribution coefficient in vadose-zone soil. K_{d_v} is a site and soil-zone specific parameter depending on the fraction organic carbon in the vadose-zone or:

$$K_{d_v} = K_{oc} \times f_{oc_v}$$

K_{d_q} : Distribution Coefficient in the Ground-Water Zone

The units used for K_{d_q} are $\frac{mg/kg \text{ (dry aquifer material)}}{mg/kg \text{ (water)}}$ and K_{d_q} is therefore unitless.

Estimation Method

This is a site-specific parameter and depends on the fraction organic carbon in the ground-water zone and on the value of K_{oc} . K_{d_q} is the product of the soil organic carbon partition coefficient (K_{oc}) and the fraction organic carbon in the ground-water zone (f_{oc_q}) (Karickhoff, 1981).

$$\begin{split} K_{d_q} &= K_{oc} \times f_{oc_q} \\ f_{oc_q} &= \frac{kg \ organic \ carbon \ (dry \ aquifer \ material)}{kg \ (solid)} \end{split}$$

Based on the estimation reported above, we obtain the following equation for the distribution coefficient in the ground-water zone. K_{d_q} is a site and soilzone specific parameter depending on the fraction organic carbon in the ground-water zone or:

$$K_{d_q} = K_{oc} \times f_{oc_q}$$

$K_{d_d} \colon$ Distribution Coefficient in Sediment Particles

The units used for K_{d_d} are $\frac{mg/kg \text{ (dry surface-water sediment)}}{mg/kg \text{ (water)}}$ and K_{d_d} is therefore unitless.

Estimation Method

This is a site specific parameter and depends on the fraction organic carbon in the surface-water sediment and the value of K_{oc} . K_{d_d} is the product of the soil organic carbon partition coefficient (K_{oc}) and the fraction of organic carbon in surface-water sediment (f_{oc_d}) [Karickhoff, 1981].

$$\begin{split} K_{d_d} &= K_{oc} \times f_{oc_d} \\ f_{oc_d} &= \frac{kg \ organic \ carbon \ (dry \ surface-water \ sediment)}{kg \ (soil)} \end{split}$$

Based on the estimation reported above, we obtain the following equation for the distribution coefficient in surface-water sediment particles. K_{d_d} is a site and soil-zone specific parameter depending on the fraction organic carbon in surface-water sediment or:

$$K_{d_d} = K_{oc} \times f_{oc_d}$$

K_{ps}: Partition Coefficient for Plant-Tissue (Above Ground Fresh Mass) Relative to Soil Concentration (Fresh Soil)

The units used for K_{ps} are $\frac{mg/kg \text{ (plant fresh mass [pFM])}}{mg/kg \text{ (soil fresh mass [sFM])}} \text{ (ppm [pFM]/ppm [sFM])}$.

No reported measurements of K_{ps} for TCE are available in the current literature. An estimation method for this parameter is thus applied.

Estimation Method

Based on a review of reported measurements of bioconcentration for 29 persistent organochlorines in plants, Travis and Arms (1988) have correlated plant-soil bioconcentration (on a dry-mass basis) in above-ground plant parts with octanol-water partition coefficients. This bioconcentration factor, $B_{\rm v}$, on a dry-weight basis is expressed as:

$$\log B_v = 1.58 - 0.58 \log K_{OW} \pm 0.73 (n=29, r^2=0.525)$$

We calculated the error term, \pm 0.73, from the mean square error of the estimator for this regression from the data provided by Travis and Arms (1988). When adjusted to a fresh-mass basis (assuming that the plant dry-mass fraction equals 0.2), this estimation equation gives the plant-soil partition coefficient, K_{ps} , expressing the ratio of contaminant concentration in mg/kg in above-ground plant fresh mass relative to contaminant concentration in mg/kg (dry soil) in the root-zone as:

$$K_{ps} = 7.7 K_{ow}^{-0.58} (CV = 4.0) ppm (pFM)/ppm (sDM)$$

Expressing the ratio of contaminant concentration in mg/kg in above-ground plant fresh mass relative to contaminant concentration in mg/kg (fresh soil), and assuming fresh soil 10% by mass water in the root-zone soil, the K_{ps} estimation is:

$$K_{ps}$$
 (est) = 7.0 $K_{ow}^{-0.58}$
 K_{ow} = 320

$$K_{ps}$$
 (est) = 0.25 ppm (pFM)/ppm (sFM)

The estimation error reported above corresponds to a CV of 4.

From the estimation method identified above, we obtain the following statistics for the partition coefficient in plant leaves relative to contaminant concentration in soil of TCE:

Arithmetic mean (coefficient of variation):
$$K_{ps} = 0.25$$
 (4) ppm (pFM)/ppm (sFM)

Kpa: Biotransfer Factors For Plant Leaves Relative to Contaminant Air Concentration

The units used for
$$K_{pa}$$
 are $\frac{mg/kg \text{ (plant fresh mass [pFM])}}{mg/\text{cubic meter of air (m}^3 \text{ [air])}} \text{ (m}^3 \text{ [a]/kg [pFM])}$

No reported measurements of K_{pa} for TCE are available in the current literature. An estimation method for this parameter is thus applied.

Estimation Method

Based on the model of Riederer (1990) for foliar uptake of gas-phase contaminants (mg/m^3) relative to contaminant concentration in plant leaves (mg/kg fresh mass), we estimate a steady-state plant-air coefficient as:

$$K_{pa} \ (m^3 \ [a]/kg \ [pFM]) = [0.5 + ((0.4 + 0.01 \times K_{ow})(RT/H))] \times 10^{-3} \ kg/m^3$$

$$R = 8.313 \ Pa-m^3/mol-K$$

$$T = 298 \ K$$

$$H = 890 \ Pa-m^3/mol$$

$$K_{ow} = 320$$

$$K_{pa}$$
 (est) = 0.011 m³ [a]/kg [pFM]

McKone (1993) has estimated that the CV associated with this partition estimation model is on the order of 14.

From the estimation method identified above, we obtain the following statistics for the partition coefficient in plant leaves relative to contaminant concentration in air for TCE:

Arithmetic mean (coefficient of variation): $K_{pa} = 0.011 (14) \text{ m}^3 [a]/\text{kg [pFM]}$

BIOTRANSFER FACTORS FOR FOOD PRODUCTS

The biotransfer factors B_t , B_k and B_e are the steady-state contaminant concentrations in, respectively; fresh meat, milk, and eggs; divided by the animals daily contaminant intake, and are expressed in units of (mg/kg)/(mg/d) or kg/d.

B_k: Steady-State Biotransfer Factors for Whole Milk Relative to Contaminant Intake by Cattle

The units used for B_k are days/kg (milk) (d/kg [milk]).

No reported measurements of B_k are available in the current literature. Estimation methods are therefore considered.

Estimation Method 1

Based on a review of biotransfer factors for 28 organic chemicals in milk, Travis and Arms (1988) developed the following geometric-mean regressions for B_{k1} based on the octanol-water partition coefficient, K_{ow} ,

$$log B_{k1} = log K_{ow} - 8.1 \pm 0.84 \ (n = 28, r^2 = 0.55)$$

Using the data provided by Travis and Arms (1988), we calculated the error term, \pm 0.84, from the mean square error of the estimator for this regression. This estimation error corresponds to a CV of 6. From the above expression and log K_{ow} of 2.51, we obtain the following statistics for the B_{k1} of TCE:

$$B_{k1}$$
 (est) = 2.56 × 10⁻⁶ days/kg (milk)
CV = 6

Estimation Method 2

The transfer of organic chemicals from animal feed to milk has also been expressed in terms of the fat-diet partition coefficient, $K_{\rm fd}$, which is the steady-state ratio of contaminant concentration in animal fat (or lipid) to contaminant concentration in animal feed with units kg (feed)/kg (fat). Kenaga (1980) reviewed cattle-dietary feeding studies for 23 chemicals, and from these studies derived the following fat-diet equation relating $K_{\rm fd}$ to $K_{\rm ow}$,

$$\log K_{fd} = 0.5 \log K_{ow} - 3.457 \pm 1 \ (n = 23, r^2 = 0.62)$$

The estimation error in this expression, \pm 1, was calculated by Kenaga (1980). From the above expression with log K_{ow} of 2.51, an assumed pasture intake by dairy cattle of 85 kg/d (McKone and Ryan, 1989), and an assumed fat content of 0.04 in milk; we obtain the following statistics for the B_{k2} of TCE:

$$B_{k2}$$
 (est)= 2.95 \times 10 $^{\text{-}6}$ days/kg (milk) CV = 14

The above estimation error corresponds to assumed CV of 14.

The estimation values reported above yield the arithmetic mean and CV reported below:

$$B_k$$
 (avg) = 2.8×10^{-6} days/kg (milk) $CV = 11$

Based on the estimation equation and the estimation error reported above, we obtain the following value for the estimated steady-state biotransfer factor for milk relative to dietary contaminant intake by dairy cattle for TCE:

Arithmetic mean (coefficient of variation):
$$B_k = 2.8 \times 10^{-6}$$
 (11) days/kg (milk)

B_t : Steady-State Biotransfer Factor for Meat Relative to Contaminant Intake by Cattle The units used for B_t are days/kg (meat) (d/kg [meat]).

No reported measurements of cattle-meat biotransfer for TCE are available in the current literature. Estimation methods are therefore considered.

Estimation Method 1

Based on a review of biotransfer factors for 36 chemicals in meat, Travis and Arms (1988) developed the following geometric-mean regression for B_{t1} based on the octanol-water partition coefficient, K_{ow} ,

$$\log B_{t1} = \log K_{OW} - 7.6 \pm 0.95 \ (n = 36, r^2 = 0.67)$$

Using the data provided by Travis and Arms (1988), we calculated the error term, ± 0.95 from the mean square error of the estimator for this regression. This estimation errors corresponds to a CV of 11. From the above expression and a log K_{ow} equal to 2.51, we obtain the following estimation:

$$B_{t1}$$
 (est) = 8.10 × 10⁻⁶ days/kg (meat) $CV = 11$

Estimation Method 2

The transfer of organic chemicals from animal feed to meat has also been expressed in terms of the fat-diet partition coefficient, $K_{\rm fd}$, which is the steady-state ratio of contaminant concentration in animal fat (or lipid) to contaminant concentration in animal feed with units kg (feed)/kg (fat). Kenaga (1980) reviewed cattle-dietary feeding studies for 23 chemicals, and from these studies derived the following fat-diet equation relating $K_{\rm fd}$ to $K_{\rm ow}$:

$$\log K_{fd} = 0.5 \log K_{ow} - 3.457 \pm 1 \ (n = 23, r^2 = 0.62)$$

The estimation error in this expression, \pm 1, was calculated by Kenaga (1980). From the above expression with log K_{ow} equal to 2.51, an assumed pasture intake by beef cattle of 60 kg/d (McKone and Ryan, 1989), and an assumed fat content of 0.4 in meat; we obtain the following estimation:

$$B_{t2} \text{ (est)} = 4.18 \times 10^{-5} \text{ days/kg (meat)}$$

$$CV = 14$$

The above estimation error corresponds to a CV of 14.

The estimation values reported above yield the arithmetic mean and CV reported below:

$$B_t \; (avg) = 2.5 \times 10^{-5} \; days/kg \; (meat)$$

$$CV = 13$$

Based on the estimation equation and the estimation error reported above, we obtain the following value for the estimated steady-state biotransfer factor for meat relative to dietary contaminant intake by cattle for TCE:

Arithmetic mean (coefficient of variation):
$$B_t = 2.5 \times 10^{-5}$$
 (13) days/kg (meat)

B_e: Steady-State Biotransfer Factors for Eggs Relative to Dietary Contaminant Intake by Chickens

The units used for B_e are days/kg (eggs) (d/kg [eggs]).

No reported measurements of egg-diet biotransfer for TCE are available in the current literature. An estimation method is therefore considered.

Estimation Method

Based on measurements of polychlorodibenzodioxins (PCDDs) and polychlorodibenzo-furans (PCDFs) concentrations in soil versus concentrations in egg-fat and adipose tissue of foraging chickens, Stephens et al. (1990) have shown that contaminant concentrations in animal fat correlate with soil concentrations. In addition, they found the fat-soil partition factor in chicken fat is roughly six times higher than the fat-soil partition factor in cattle. However, the fraction of total intake represented by soil in the chicken feed is higher than in the cattle feed. Based on these observation and what is discussed in the above B_k and B_t sections, we (a) assume that the fat-diet partition factor in chickens is similar to that in cattle, (b) use log K_{fd} = log K_{ow} - 4.9 to estimate the K_{fd} for chickens, and (c) use the fat content of eggs (0.08) and feed intake of chickens (0.12 kg/d [fresh mass]) to obtain the following estimate of a biotransfer factor, B_e , from chicken feed to eggs with units d/kg (eggs):

$$\log B_e = \log K_{OW} - 5.1$$
$$\log K_{OW} = 2.51$$

$$B_e = 2.6 \times 10^{-3} \text{ d/kg (eggs)}$$

We estimate the CV in this expression is 14.

Based on the estimation equation and the estimation error reported above, we obtain the following value for the estimated steady-state biotransfer factors for eggs relative to dietary contaminant intake by chickens for TCE:

Arithmetic mean (coefficient of variation):

$$B_e = 2.6 \times 10^{-3}$$
 (14) days/kg (eggs)

B_{bmk}: Biotransfer Factor for Human Breast Milk Relative to Dietary Contaminant Intake by the Mother

The units used for B_{bmk} are days/kg (mothers milk) (d/kg [mothers milk]).

Estimation Method

No experimental results quantifying B_{bmk} are available in the current literature, an estimation method (Smith, 1987), using a K_{ow} for TCE of 320, is thus applied:

$$B_{bmk} = 2 \times 10^{-7} K_{ow}$$
$$K_{ow} = 320$$

 $B_{bmk} = 6.4 \times 10^{-5} \text{ days/kg (mothers milk)}$

The CV of the above estimation method is approximately 10.

Based on the estimation equation and the estimation error reported above, we obtain the following value for the estimated biotransfer factor for human breast milk concentration relative to dietary contaminant intake by the mother for TCE:

Arithmetic mean (coefficient of variation):
$$B_{bmk} = 6.4 \times 10^{-5}$$
 (10) days/kg (mothers milk)

BCF: Bioconcentration Factors for Fish Relative to Water Concentration

The units used for BCF (fish/water) are $\frac{mg/kg \text{ (fish)}}{mg/liter \text{ (water)}}$, and BCF is therefore unitless.

Experimental Values:

reported at 16 °C as a log BCF (fish/water) of 1.23 by Veith et al. (1980) using bluegill sunfish (*Lepomis machrochirus*) and a concentration of 8.2 μg/l [also reported by Mackay et al. (1993)]

90 reported as a fish BCF of 90 by Freitag et al. (1985) using Golden Ide (Idus idus melanotus) for 3 days

From the 2 measured values* reported above, we obtain the following mean and approximate CV for BCF in fish relative to contaminant concentration in water for TCE:

Arithmetic mean (coefficient of variation): BCF (fish/water) = 53 (1)

Range: 17 to 90

Estimation Method

For fish, the BCF is taken as the ratio of concentration of a xenobiotic substance in fish flesh (or lipids) to the contaminant's concentration in water (Mackay, 1982) The BCF for neutral organic compounds can be estimated from regression equations based on selected physicochemical properties, particularly a compound's K_{ow} or aqueous solubility. Mackay (1982) recommends:

$$BCF = 0.048 \text{ K}_{oW}$$

$$K_{oW} = 320$$

$$BCF \text{ (est)} = 15$$

$$CV = 0.6$$

The reported GSD is 1.8 which corresponds to an estimation error of 0.6.

$\mathbf{K}_{\mathbf{p}_{-}\mathbf{w}}$: Human Skin Permeability Coefficient Relative to Contaminant Concentration in Water

The units used for K_{p_w} are centimeters/hour (cm/hr).

No reported measurements of K_{p_w} for TCE are available in the current literature. An estimation method for this parameter is thus applied.

Estimation Method

Because dermal transfer is considered a nonsteady-state event, diffusion models require input parameters which are difficult to measure, such as the stratum corneum diffusion coefficient (D_{sc}) [Flynn and Amidon, 1991]. Estimation of aqueous biotransfer of TCE is calculated with the following equation based on the estimation method of McKone and Howd (1992).

$$K_{p_W} = MW^{\text{-}0.6} \quad 0.33 + \frac{0.0025}{2.4 \times 10^{\text{-}6} + 3 \times 10^{\text{-}5} \ K_{oW}^{0.8}} \, ^{\text{-}1}$$

$$K_{ow} = 320$$

 $MW = 131.4 \text{ g/mol}$

$$K_{p_w} = 0.047 \text{ cm/hr}$$

who report a coefficient of variation equal to 2.4

Based on the estimation equation and the estimation error reported above, we obtain the following value for the estimated human skin permeability coefficient relative to contaminant water concentration for TCE:

Arithmetic mean (coefficient of variation):
$$K_{p_w} = 0.047 (2.4) \text{ cm/hr}$$

K_m: Partition Coefficient for Human Skin Relative to Contaminant Concentration in Water or Soil

The units used for K_m are $\frac{mg/kg \text{ (skin)}}{mg/liter \text{ (water)}} \text{ (ppm [skin]/ppm [water])}$.

No experimental values for K_m are currently available in the literature, therefore an estimation method is considered.

Estimation Method

Experimental values quantifying dermal transfer of TCE in water, or for water in a soil matrix, may depend on pH, particle size and organic carbon content (Flynn and Amidon, 1991). An estimation method based on McKone and Howd (1992) is therefore used here.

$$K_{\rm m} = 0.64 + (0.25 \text{ K}_{\rm ow}^{0.8})$$

 $K_{\rm ow} = 320$

$$K_m = 26 ppm (skin)/ppm (water)$$

The reported geometric standard deviation of 1.3 in this estimation method corresponds to a CV of 0.27.

Based on the estimation equation and the estimation error reported above, we obtain the following value for the partition coefficient into human skin relative to TCE water or soil concentration:

Arithmetic mean (coefficient of variation): $K_m=26 (0.27) \text{ ppm (skin)/ppm (water)}$

Thalf a: Reaction Half-Life in Air

The units used for T_{half} a are days.

Reported Values

2.8	reported at 23 °C as a reaction rate constant (k_{OH}) of 2.86 \times 10 ⁻¹²
	cm ³ /molecule-sec by Edney (1986) using an hydroxyl radical (OH)
	concentration in the trophosphere as 1×10^6 molecules/cm ³

- 3.38 reported at 23 °C as a k_{OH} of 2.37 \times 10⁻¹² cm³/molecule-sec by Chang and Kauffman (1977) using halocarbon concentrations from 0.15 \times 10¹⁴ to 1.56 \times 10¹⁴ molecules /cm³ and assuming a (OH) concentration of 1 \times 10⁶ molecules /cm³
- 3.41 reported at 25 °C as a k_{OH} of 2.35 \times 10⁻¹² cm³/molecule-sec by Davis et al. (1975) using 100 torr of helium and assuming a (OH) concentration of 1×10^6 molecules /cm³
- 3.49 reported as a k_{OH} of 2.3 \times 10⁻¹² cm³/molecule-sec in the trophosphere by Atkinson et al. (1984) assuming a (OH) concentration of 1 \times 10⁶ molecules/cm³
- 3.65 reported as a k_{OH} of 2.2 \times 10^{-12} $cm^3/molecule\text{-sec}$ by Singh et al. (1981) using field air sample collections and an average atmospheric OH concentration of 1 \times 10^6 molecules/cm^3
- 4.01 reported as a k_{OH} of 2 \times 10⁻¹² cm³/molecule sec by Howard (1976) using laser magnetic resonance detection in a discharge-flow system and assuming an average atmospheric (OH) concentration of 1 \times 10⁶ molecules/cm³

From the 6 experimental values reported above, we obtain the following statistics on the reaction half-life for TCE in air:

Arithmetic mean (coefficient of variation): $T_{half_a} = 3.5 (0.11) days$

Range: 2.8 to 4.0 days

Thalf g: Reaction Half-Life in Ground-Surface Soil

The units used for $T_{half\ g}$ are days.

Reported Values

33 to reported as an estimated half-life in soil of 33 to 230 days by Olsen and

230 Davis (1990)

reported as an estimated half-life in soil of 730 days by Jury (1990)

From the 3 measured values above, and the assumption that the actual range of values may be a factor of 5 higher or lower than this range, we obtain the following statistics for the reaction half-life for TCE in surface soil:

Arithmetic mean (coefficient of variation):

 $T_{half\ g} = 930\ (1.7)\ days$

Range: 7 to 3650 days

Thalf s: Reaction Half-Life in Root-Zone Soil

The units used for Thalf s are days.

Reported Values

reported as an estimated half-life in soil of 33 to 230 days by Olsen and

230 Davis (1990)

reported as an estimated half-life in soil of 730 days by Jury (1990)

From the 3 measured values above, and the assumption that the actual range of values may be a factor of 5 higher or lower than this range, we obtain the following statistics for the reaction half-life for TCE in root-zone soil:

Arithmetic mean (coefficient of variation):

 $T_{half s} = 930 (1.7) days$

Range: 7 to 3650 days

Thalf v: Reaction Half-Life in Vadose-Zone Soil

The units used for $T_{half\ v}$ are days.

Reported Values

33 to reported as an estimated half-life in soil of 33 to 230 days by Olsen and

230 Davis (1990)

260	reported at 17 °C as a half-life corresponding to 260 days by Wilson et al. (1986) using ground water samples (pH = 7.3; OC = 344 mg/L) incubated in a dark anaerobic microcosm for 120 weeks and assuming first order degradation
402	reported at 17 °C as a degradation half-life corresponding to 402 days by Wilson et al. (1983a) using aquifers samples (2.1-3.6 m) in Oklahoma and Louisiana (pH = 4.2-5.7; f_{oc} =1.4-2%) at concentrations of 600-800 µg/L and assuming first-order degradation
730	reported as an estimated half-life in soil of 730 days by Jury (1990)
2888	reported as a biotransformation rate constant of 1 \times 10 ⁻⁵ hrs. $^{-1}$ by Silka (1988) using data from measurements at a sand and gravel outwash aquifer located in Tacoma, WA

From the 6 values above, we obtain the following statistics for the reaction half-life for TCE in vadose-zone soil:

Arithmetic mean (coefficient of variation): $T_{half_v} = 760 (1.4) days$

Range: 33 to 2888 days

T_{half_q} : Reaction Half-Life in Groundwater

The units used for T_{half_q} are days.

Reported Values

128

	(1989) using an anoxic methanogenic nutrient solution (autoclaved) in the dark and a 25% TCE solution
260	reported at 17 °C as a half-life corresponding to 260 days by Wilson et al. (1986) using ground water samples (pH = 7.3; OC = 344 mg/L) incubated in a dark anaerobic microcosm for 120 weeks and assuming first order degradation
325	reported as a decomposition half-life corresponding to 325 days by Dilling (1975) in sealed flasks without light for one year and assuming first order kinetics

reported as a degradation half-life of 128 days by Baek and Jaffe

reported at 17 °C as a degradation half-life corresponding to 402 days by Wilson et al. (1983a) using aquifers samples (2.1-3.6 m) in Oklahoma and Louisiana (pH = 4.2-5.7; f_{oc} =1.4-2%) at concentrations of 600-800 µg/l and assuming first-order degradation

2888

reported as a biotransformation rate constant of 1×10^{-5} hrs. ⁻¹ by Silka (1988) using data from measurements at a sand and gravel outwash aquifer located in Tacoma, WA

From the 6 values above, we obtain the following statistics for the reaction half-life for TCE in groundwater:

Arithmetic mean (coefficient of variation):

 $T_{half q} = 800 (1.5) days$

Range: 128 to 2888 days

Thalf w: Reaction Half-Life in Surface Water

The units used for T_{half_w} are days.

Reported Values

185 to 325

7	reported as a degradation half-life of 7 days by Tabak et al. (1981) in a municipal wastewater mixture due to aerobic biodegradation
13.4	reported at 12 °C as a degradation half-life corresponding to 13.4 days by Jensen and Rosenberg (1975) using 0.1 to 1ppm in a closed seawater system, with and without sunlight, for 14 days and assuming first order degradation
31 to 90	reported as a first order degradation half-life of 31 to 90 days reported by Zoetemann et al. (1980) and based on the concentration reduction between sample points on the Rhine River and a lake in the Rhine basin [also cited in Mackay et al. (1993)]
78 to 128	reported as a degradation half-life of 78 to 128 days by Baek and Jaffe (1989) in an anoxic methanogenic nutrient solution (autoclaved or bacterially active) in the dark at a 25% TCE solution

reported as a decomposition half-life corresponding to 185 to 325 days by

Dilling (1975) in sealed flasks with and without light for one year and

From the 8 values above, we obtain the following statistics for the half-life for TCE in surface water:

Arithmetic mean (coefficient of variation):

 $T_{half_w} = 120 (0.88) days$

assuming first order kinetics

Range: 7 to 325 days

Thalf d: Reaction Half-Life in Surface Water Sediment

The units used for T_{half} d are days.

Reported Values

78 to 128	reported as a degradation half-life of 78 to 128 days by Baek and Jaffe (1989) using an anoxic methanogenic nutrient solution (autoclaved and bacterially active) in the dark and a 25% TCE solution
260	reported at 17 °C as a half-life corresponding to 260 days by Wilson et al. (1986) using ground water samples (pH = 7.3 ; OC = 344 mg/L) incubated in a dark methanogenic microcosm for 120 weeks and assuming first order degradation
402	reported at 17 °C as a degradation half-life corresponding to 402 days by Wilson et al. (1983a) using aquifers samples (2.1-3.6 m) in Oklahoma and Louisiana (pH = 4.2-5.7; f_{oc} =1.4-2%) at concentrations of 600-800 µg/l and assuming first-order degradation

From the 7 values above, we obtain the following statistics for the half-life of TCE in sediment:

> Arithmetic mean (coefficient of variation): $T_{half\ d} = 220 (0.67) days$

Range: 78 to 402 days

References

Abdul, S. A., Gibson, T. L., and D. N. Rai, 1987. Statistical Correlations for Predicting the Partition Coefficient for Nonpolar Organic Contaminants Between Aquifer Organic Carbon and Water, Hazardous Waste and Hazardous Materials 4(3):211-222

Ashworth, R.A., et al. 1986. Air-Water Partitioning Coefficients of Organics in Dilute **Aqueous Solutions**, paper presented at the American Institute of Chemical Engineers 1986 National Meeting; Boston, MA

Ashworth, R. A., Howe, G. B., Mullins, M. E., and Rogers, T. N., 1988. Air-Water Partitioning Coefficients of Organics in Dilute Aqueous Solutions, J. Haz. Mater. 18:25-36

Atchison, J. and J.A.C. Brown, 1957. The Lognormal Distribution, Cambridge University; New York, NY. pp 125

Atkinson, R. and W. P. Carter, 1984. Kinetics and Mechanisms of the Gas-Phase Reactions of Ozone with Organic Compounds under Atmospheric Conditions, Chem Rev. 84:437-470

- Atkinson, R., 1985. Kinetics and Mechanisms of the Gas Phase Reaction of Hydroxyl Radical with Organic Compounds Under Atmospheric Conditions, Chem. Rev. 85: 69–201
- Baek, N. H. and P. R. Jaffe, 1989. Anaerobic Mineralization of Trichloroethylene. International Conference on Physiochemical and Biological Detoxification of Hazardous Wastes: May 3-5, 1988; Atlantic City, NJ. Edited by Y. C. Yu; Lancaster, Pa.: Technomic
- Banerjee, S., S.H. Yalkowsky, and S.C. Valvani 1980. Water Solubility and Octanol/Water Partition Coefficient of Organics. Limitations of Solubility-Partition Coefficient Correlation, *Environ. Sci. Technol.* 14(10):1227-1229
- Barrio-Lage, G., F.Z. Parsons, R.S. Nassar and P.A. Lorenzo, 1986. Sequential Dehalogenation of Chlorinated Ethenes, Environ. Sci. Technol. 20:96-99
- **Boublik, T., V. Fried, and E. Hála, 1984.** The Vapour Pressures of Pure Substances: Selected Values of the Temperature Dependence of the Vapour Pressures of Some Pure Substances in the Normal and Low Pressure Region (Second Revised Edition), Physical Sciences Data; 17 Elsevier Science; Amsterdam, The Netherlands
- Bouwer, E.J., B. Rittman, and P.L. McCarty, 1981. Anaerobic Degradation of 1- and 2-Carbon Halogenated Aliphatic Organic Compounds, Environ. Sci. Technol. 15:596–599
- Broholm, K., J.A. Cherry and S. Feenstra, 1992. Dissolution of Heterogeneously Distributed Solvents Residuals. Proceedings of Subsurface Restoration Conference, Third International Conference on Ground Water Quality Research, June 21-24, 1992, Dallas, TX. National Center for Ground Water Research, Rice University, Environ. Sci. & Engineering., P.O. Box 1892, Houston TX 77251, pp 96-98
- Budavari, S., O'Neil, M. J., Smith, A. and Heckelman, P. E. Eds., 1989. The Merck Index, Eleventh Ed., Merck
- Chang, J.S., and F. Kauffman, 1977. Kinetics of the Reactions of Hydroxyl Radicals with Some Halocarbons: CHFCl₂, CHF₂Cl, CH₃CCl₃, C2HCl₃, and C₂Cl₄, *J. Chem. Phys.* 11:4989-4994
- Chiou, C. T., and V. H. Freed, 1977. Chemodynamic Studies on Bench Mark Industrial Chemicals (Annual Report), Oregon State University Dept. of Agricultural Chemistry; Corvallis, OR for Research Applied to National Needs, National Science Foundation; Washington, DC. NTIS PB-274-263, NSF/RA-770286
- Clayton, G. D. and Clayton, F. E. Ed., 1981. Patty's Industrial Hygiene & Toxicology. Third ed., John Wiley & Sons; New York, NY., Vol. 2B:3491-3497
- Coca, J. and R. Diaz, 1980. J. Chem. Eng. Data, 25(1):80-83
- Daubert, T. E. and R. P. Danner, 1985. Data Compilation of Pure Compounds, Vol. I, July 1985. Design Institute for Physical Property Data American Institute of Chemical Engineers; 345 East 47th Street, New York, NY
- Daubert, T. E. and R. P. Danner, 1989. Data Compilation of Pure Compounds, Vol. II, 1989. Design Institute for Physical Property Data American Institute of Chemical Engineers; 345 East 47th Street, New York, NY

Davis, D.D., W. Bollinger and S. Fischer, 1975. A Kinetics Study of the Reaction of the OH Free Radical with Aromatic Compounds. I. Absolute Rate Constants for Reaction with Benzene and Toluene at 300 degrees K, J. Physic. Chem. 79(3):293-294

Dean, J.D., Ed. 1985. Lange's Handbook of Chemistry, McGraw-Hill; New York, NY. 13th ed

Dilling, W.L., N.B. Tefertiller and G.J. Kallos, 1975. Evaporation Rates and Reactivities of Methylene Chloride, Chloroform, 1,1,1-Trichloroethane, Trichloroethylene, Tetrachloroethylene, and Other Chlorinated Compounds in Dilute Aqueous Solutions, Environ. Sci. Technol. 9(9):833-838

Doust, H.G., and J.C. Huang, 1992. The Fate and Transport of Hazardous Chemicals in the Subsurface Environment, Water Sci. Technol. 25(1):169-176

Dreisbach, R., 1959. Advances in Chemistry Series (22): Physical Properties of Chemical Compounds--II, American Chemical Society; Washington, DC407

Edney, E.O., T.E. Kleindienst and E.W. Corse, 1986. Room Temperature Rate Constants for the Reaction of OH with Selected Chlorinated and Oxygenated Hydrocarbons, *Int. J. Chem. Kinet.* 18:1355-1371

Flynn and Amidon, 1991. Dermal Exposure Assessment: Principles and Applications, Interim Report. USEPA Jan, 1992. Office of Research and Development; Washington, DC., EPA/600/8-91/011B

Freitag, D., L. Ballhorn, H. Geyer and F. Korte, 1985. Environmental Hazard Profile of Organic Chemicals, *Chemosphere* 14(10):1589-1616

Friesel, P., G. Milde and B. Steiner, 1984. Interactions of Halogenated Hydrocarbons with Soils, Fresenius Z. Anal. Chem. 319:160-164

Fuller, E.N., P.D. Schettler, and J.C. Giddings, 1966. A New Method for Prediction of Binary Gas-Phase Diffusion Coefficients, *Ind. Eng. Chem.* 58:19-27

Garbarini, D.R., and L.W. Lion, 1985. Evaluation of Sorptive Partitioning of Nonionic Pollutants in Closed Systems by Headspace Analysis, *Environ. Sci. Technol.* 19(11):1122-1128

Garbarini, D.R., and L.W. Lion, 1986. Influence of the Nature of Soil Organics on the Sorption of Toluene and Trichloroethylene, Environ. Sci. Technol. 20(12):1263-1269

Glave and C. Hansch, 1985. Unpublished Results in the Log P and Related Parameters Database, created and updated by the Medicinal Chemistry Project at Pomona College, Claremont, CA, under the direction of Corwin Hansch and Albert Leo — 1987 Printout

Gossett, J.M. 1987. Measurement of Henry's Law Constants for C₁ and C₂ Chlorinated Hydrocarbons, Environ. Sci. Technol. 21, 202–208

Grathwohl, P. 1990. Influence of Organic Matter From Soils and Sediments From Various Origins on the Sorption of Some Chlorinated Aliphatic Hydrocarbons: Implications on K_{oc} Correlations, Environ. Sci. Technol. 24:1687–1693

Hamburg, M. 1970. Statistical Analysis for Decision Making. Harcourt Brace Jovanovich; San Diego, CA

Hansch, C., and A. Leo 1987. The Log P and Related Parameters Database, created and updated by the Medicinal Chemistry Project at Pomona College; Claremont, CA, under the direction of Corwin Hansch and Albert Leo — 1987 Printout

Hansch, C., and A. Leo, 1985. The Log P and Related Parameters Database, created and updated by the Medicinal Chemistry Project at Pomona College under the direction of Corwin Hansch and Albert Leo — 1985 Printout, Medchem Project Issue No. 26, Pomona College; Claremont, CA

Hansch, C., and D. Elkins, 1971. Chem. Rev. 71:525

Harnisch, M., H.J. Mockel and G. Schulze, 1983. Relationship Between Log P_{ow} Shake-Flask Values and Capacity Factors Derived from Reversed-Phase High-Performance Liquid Chromatography for N-Alkylbenzenes and Some OECD Reference Substances, *J. Chromatog.* 282:315-332

Hendry and Kenley, 1976. Atmospheric Reaction Products of Organic Compounds, EPA Report. EPA-560/12-79-001, pp 80

Hertz, W and W. Rathmann, 1912. Chem.-Ztg. 36:1417

Holden, N. E., 1980. Pure Appl. Chem. 52:3249

Horvath, A. L., 1982. Halogenated Hydrocarbons, Solubility–Miscibility with Water Marcel Dekker; New York, NY

Howard, C.J., 1976. Rate Constants for the Gas-Phase Reactions of OH Radicals with Ethylene and Halogenated Ethylene Compounds, J. Chem. Phys. 65(11):4771-4777

Howard, P. H., R. S. Boethling, W. F. Jarvis, W. M. Meylan, and E. M. Michalenko, Eds. 1991. *Handbook of Environmental Degradation Rates*, Lewis; Chelsea, MI

Howard, P.H., G.W. Sage, W.F. Jarvis, and D.A. Gray, Eds. 1990. Handbook of Environmental Fate and Exposure Data for Organic Chemicals. Volume II. Solvents Lewis; Chelsea, MI

International Programme on Chemical Safety, 1984. World Health Organization Environmental Health Criteria 50: Trichloroethylene, Geneva, Switz.

Jensen, S., and R. Rosenberg, 1975. Degradability of Some Chlorinated Aliphatic Hydrocarbons in Seawater and Sterilized Water, *Water Res.* 9:659–661

Jury, W. A, D. Russo, G. Streile and H. El Abd, 1990. Evaluation of Volatilization by Organic Chemicals Residing Below the Soil Surface, Water Resour. Res. 26(1):13-20

Karickhoff, S. W., 1981. Semi-Empirical Estimation of Sorption of Hydrophobic Pollutants of Natural Water Sediments and Soils, Chemosphere 10(8):833-846

Kenaga, E. E., 1980. Correlation of Bioconcentration Factors of Chemicals in Aquatic and Terrestrial Organisms with Their Physical and Chemical Properties, *Environ. Sci. Technol.* 14(5):553-556

Kenaga, E. E., and C. A. I. Goring 1980. Relationship Between Water Solubility, Soil Sorption, Octanol-Water Partitioning, and Concentration of Chemicals in Biota, in Aquatic Toxicology, ASTM STP 707, J. G. Eaton, P. R. Parrish, and A. C. Hendricks, Eds. American Society for Testing and Materials; Philadelphia, PA. pp 78–115

Kirk-Othmer, 1985. Kirk-Othmer Encyclopedia of Chemical Technology. Wiley-Interscience, John Wiley & Sons; New York, NY. 3rd ed., Vol. 5

Kirk-Othmer Encyclopedia of Chemical Technology, 1964. Chlorocarbons and Chlorohydrocarbons,, Wiley Interscience; New York, NY. Vol. 5, 2nd ed.

Lee, J.F., J.R. Crum, and S.A. Boyd 1989. Enhanced Retention of Organic Contaminants by Soils Exchanged with Organic Cations, Environ. Sci. Technol. 23:1365–1372

Lee, K., and N. Tanaka, 1990. [Method for Determining Partition Coefficient of Trichloroethylene], Suishitsu Odaku Kenkyu, 13(4):55-59 (in Japanese)

Leighton, D.T., and J.M. Calo, 1981. Distribution Coefficients of Chlorinated Hydrocarbons in Dilute Air-Water Systems for Groundwater Contamination Applications, J. Chem. Eng. Data 26:382–385

Liljestrand, H. M. and R. J. Charbeneau, 1987. Analysis and Interpretation of Batch Equilibrium and Column Studies of the Partitioning of Chlorinated Hydrocarbons to Soil Materials from the PPG Lake Charles Site. Environmental and Water Resources Engineering, Department of Civil Engineering, The University of Texas at Austin; Austin, TX 78712

Lincolff, A.H., and J.M. Gossett, 1984. "The Determination of Henry's Law Constant for Volatile Organics by Equilibrium Partitioning in Closed Systems", in *Gas Transfer at Water Surfaces*, W. Brutsaert, and G.H. Jirka, Eds. Reidel Publishing; Dordrecht, The Netherlands, pp 17–25

Lyman, W. J., W. F. Reehl, and D. H. Rosenblatt, 1990. Handbook of Chemical Property Estimation Methods: Environmental Behavior of Organic Compounds, American Chemical Society; Washington, DC

Lyman, W. J., W. F. Reehl, and D. Rosenblatt, 1982. Handbook of Chemical Property Estimation Methods: Environmental Behavior of Organic Compounds, American Chemical Society, Washington, D. C.

Mackay, D. 1982. Correlation of Bioconcentration Factors, Environ. Sci. Technol. 16:274–278

Mackay, D., W. Y. Shiu, and K. L. Ma, 1992. Illustrated Handbook of Physical Chemical Properties and Environmental Fate for Organic Chemicals. Vol. II. Polynuclear Aromatic Hydrocarbons, Polychlorinated Dioxins, and Dibenzofurans. Lewis; Boca Raton, FL

Mackay, D., W.-Y. Shiu, and K.-C. Ma, 1993. Illustrated Handbook of Physical-Chemical Properties and Environmental Fate for Organic Chemicals. Volume III. Volatile Organic Chemicals, Lewis; Boca Raton, FL

McDonald, H. J., 1944. J. Phys. Chem. 48:47

McGovern, E. W., 1943. Chlorohydrocarbon Solvents. Ind. Eng. Chem. 35(12):1230-1239

McKone, T. E., and Howd, R. A., 1992. Estimation Dermal Uptake of Nonionic Organic Chemicals from Water and Soil: I. Unified Fugacity-Based Models for Risk Assessments, *Risk Analysis* 12(4) 543-557

McKone, T. E., and Ryan, P. B., 1989. Human Exposures to Chemicals Through Food Chains: An Uncertainty Analysis, Environ. Sci. Technol. 23:1154-1163

McKone, T.E., 1993. The Precision of QSAR Methods for Estimating Intermedia Transfer Factors in Exposure Assessments, SAR and QSAR in Environmental Research, 1:41-51

Montgomery, J. H. and L. M. Welkom, 1990. Groundwater Chemicals Desk Reference. Lewis; Chelsea, MI

Morgan, D. L., Cooper, S W., Carlock, D L., Sykora, J J., Sutton, B., Mattie, DR., McDougal, JN. Dermal Absorption of Neat and Aqueous Volatile Organic Chemicals in the Fischer 344 Rat, *Environ-Res*, 55(1):51-63

Munz, C., and P.V. Roberts, 1987. Air-Water Phase Equilibria of Volatile Organic Solutes, J. Am. Water Works Assoc., (May):62–69

Mudder, T. 1981. Development of Empirical Structure-Biodegradability Relationships and Testing Protocol for Slightly Soluble and Volatile Priority Pollutants, *Diss. Abstr. Int. B.* 42:1804

Munz, C., and P.V. Roberts 1982. Technical Report No. 262, Department of Civil Engineering, Stanford University; Stanford, CA

Nathan, M. F., 1978. Chem. Eng. 85(3):93-100

Neely, W.B., 1976. Predicting the Flux of Organics Across the Air/Water Interface. Control of Hazardous Material Spills: Proceedings of 1976 National Conference on Control of Hazardous Material Spills, April 25-28, 1976, New Orleans, LA. Hazardous Materials Control Research Institute; Silver Spring, MD pp 197-200

Neely, W.B., D.R. Branson, and G.E. Blau, 1974. Partition Coefficient to Measure Bioconcentration Potential of Organic Chemicals in Fish, *Environ. Sci. Technol.* 8:1113–1115

Olsen, R.L., and A. Davis, 1990. Predicting the Fate and Transport of Organic Compounds in Groundwater: Part I, Haz. Mat. Control 3(2):39-64

Pankow, J.F., and M.E. Rosen, 1988. Determination of Volatile Compounds in Water by Purging Directly to a Capillary Column with Whole Column Cryotrapping, *Environ. Sci. Technol.* 22:398-405

Piwoni, M.D., and P. Banerjee, 1989. Sorption of Volatile Organic Solvents from Aqueous Solution onto Subsurface Solids, J. Contam. Hydrol. 4:163-179

Polak, J., Murakami, S., Lam, V. T., and G. C. Benson, 1970. J. Chem. Eng. Data 15:323

Reid, R. C., J. M. Prausnitz, and B. E. Poling, 1987. The Properties of Gases and Liquids McGraw Hill; New York, NY. 4th ed

Reiderer, M. 1990. Estimating Partitioning and Transport of Organic Chemicals in the Foliage/Atmosphere System: Discussion of a Fugacity-Based Model, Environ. Scil. Technol. 24:829–837

Riddick, J. A., Bunger, W. B., Sakano, T. K., 1986. Organic Solvents: Physical Properties and Mehods of Purification, 4th ed., John Wiley & Sons; New York, NY. Vol. II: No. 319

Riddick, J. A., and W.B. Bunger, 1970. Organic Solvents. Physical Properties and Methods of Purification, (3rd ed., Vol. 2, Wiley–Interscience; New York, NY. 603 pp

- Rogers, R.D., and J.C. McFarlane, 1981. Sorption of Carbon Tetrachloride, Ethylene Dibromide, and Trichloroethylene on Soil and Clay, Environ. Monit. Assess. 1:155-162
- Rutherford, D.W., and C.T. Chiou, 1992. Effect of Water Saturation in Soil Organic Matter on the Partition of Organic Compounds, Environ. Sci. Technol. 26:995-970
- Ryan, J.A., R.M. Bell, J.M. Davidson and G.A. O'Connor, 1988. Plant Uptake of Non-Ionic Organic Chemicals from Soils, Chemosphere 17(12):2299-2323
- Sax, N. I. and R. J. Lewis Sr., 1989. Dangerous Properties of Industrial Materials, 7th Ed. Van Nostrand Reinhold; New York, NY
- Sax, N. I. and R. J. Lewis Sr., 1987. Hawley's Condensed Chemical Dictionary. Eleventh ed., Van Nostrand Reinhold; New York, NY
- Schwarzenbach, R. P., Gschwend, P. M., and D. M. Imboden, 1993. Environmental Organic Chemistry, Wiley-Interscience, John Wiley and Sons; New York, NY
- Sconce, J. S., Ed., 1962. Chlorine: Its Manufacture, Properties, and Uses, Reinhold; New York, NY. 901 pp
- Seip, H.M., J. Alstad, G.E. Carlberg, K.Martinsen and R. Skaane, 1986. Measurement of Mobility of Organic Compounds in Soils, Sci. Total Environ. 50:87-101
- Silka, L.R., and D.A. Wallen, 1988. Observed Rates of Biotransformation of Chlorinated Aliphatics in Groundwater. Superfund '88, Proceedings of the 9th National Conference, Nov. 28-30, Hazardous Materials Control Research Institute; Washington, DC pp 138-141
- Singh, H.B., L.J. Salas, A.J. Smith and H. Shigeishi, 1981. Measurements of Some Potentially Hazardous Organic Chemicals in Urban Environments, Atmos. Environ. 15:601-612
- Smith, A. H. 1987. Infant Exposure Assessment for Breast Milk Dioxins and Furans Derived from Waste Incineration Emissions, Risk Analysis 7:347-353
- Stephens, R. D., Harnly, M., Haywood, G. G., Chang, R. R., Flattery, J., Petreas, M. X., Goldman, L., 1990. Bioaccumulation of Dioxins in Food Animals II: Controlled Exposure Studies, *Chemosphere* 20:1091-1096
- Stull, D. R., 1947. Vapor Pressure of Pure Substances: Organic Compounds, Ind. Eng. Chem. 39:517–540
- Tabak, H. H., Quave, S. A., Mashni, C. I., and Barth, E. F., 1981. Biodegradability Studies with Organic Priority Pollutant Compounds. J. Water Pollut. Control Fed., 53:153-1518
- Tancréde, M.V., and Y. Yanagisawa 1990. An Analytical Method to Determine Henry's Law Constant for Selected Volatile Organic Compounds at Concentrations and Temperatures Corresponding to Tap Water Use, J. Air Waste Manage. Assoc. 40:1658–1663
- Tewari, Y.B., M.M. Miller, S.P. Wasik and D.E. Martire, 1982. Aqueous Solubility and Octanol/Water Partition Coefficient of Organic Compounds at 25.0 Deg. C, J. Chem. Eng. Data 27:451-454
- Topp, E., I. Scheunert, A. Attar, and F. Korte, 1986. Factors Affecting the Uptake of ¹⁴C-Labeled Organic Chemicals by Plants From Soil, Ecotoxicol. Environ. Saf. 11:219–228

- Travis, C. C., and A. D. Arms, 1988. Bioconcentration of Organics in Beef, Milk, and Vegetation, Environ. Sci. Technol. 22:271–274
- Tse, G., Orbey, H., and Sandler, S. I. Infinite Dilution Activity Coefficients and Henry's Law Coefficients of Some Priority Water Pollutants Determined By a Relative Gas Chromatographic Method, *Environ. Sci. Tech.* 26(10):2017-2022
- Urano, K., and C. Murata 1985. Adsorption of Principal Chlorinated Organic Compounds on Soil, Chemosphere 14(3/4):293–299
- US Dept. of Health and Human Services Agency for Toxic Substances and Disease Registry, 1992. Toxicological Profile for Tetrachloroethylene, ATSDR/TP-92/18
- Veith, G.D., K.J. Macek, S.R. Petrocelli, and J. Caroll, 1980. An Evaluation of Using Partition Coefficients and Water Solubility to Estimate Bioconcentration Factors for Organic Chemicals in Fish, in: Aquatic Toxicology, ASTM STP 707, J. G. Eaton, P. R. Parrish, and A. C. Hendricks, Eds., American Society for Testing and Materials; Philadelphia, PA pp 117–119
- **Verschueren**, K., 1983. Handbook of Environmental Data on Organic Chemicals, Second Ed., Van Nostrand Reinhold; New York, NY
- Weast, R. C., Lide, D. R., Astle, M.J. and Beyer, W.H. Eds. 1989. CRC Handbook of Chemistry and Physics: A Ready Reference Book of Chemical and Physical Data, 67th ed. CRC; Boca Raton, FL
- Weast, R. C., M.J. Astle, and W.H. Beyer, Eds. 1986. CRC Handbook of Chemistry and Physics: A Ready Reference Book of Chemical and Physical Data, 67th Edition, CRC; Boca Raton, FL
- Warner, H.P., J.M. Cohen and J.C. Ireland, 1987. Determination of Henry's Law Constants of Selected Priority Pollutants., EPA/600/D-87/229 U.S. EPA, Cincinnati, OH. NTIS: PB87-212684 14
- Wilke, C.R., and P. Chang, 1955. Correlation of Diffusion Coefficients in Dilute Solutions, AIChE J. 1:264–270
- Wilson, B. H., Smith G. B., and J. F., Rees, 1986. Biotransformations of Selected Alkylbenzenes and Halogenated Aliphatic Hydrocarbons in Methanogenic Aquifer Material: A Microcosm Study, *Environ. Sci. Technol.* 20(10):997-1002
- World Health Organization International Agency for Research on Cancer, 1979. IARC Monographs on the Evaluation of the Carcinogenic Risk to Chemicals to Humans, Trichloroethylene, Vol. 20
- Wilson, J. T., McNabb, J. F., Wilson, B. H., and M. J. Noonan, 1983a. Biotransformation of Selected Organic Pollutants in Ground Water, Developments in Industrial Microbiology, Vol. 24, Proceedings of the Thirty-Ninth General Meeting of the Society for Industrial Microbiology, St. Paul, MN. Aug. 1982
- Windholz, M., S. Budavari, R. F. Blumetti, and E. S. Otterbein, Eds., 1983. The Merck Index: An Encyclopedia of Chemicals, Drugs, and Biologicals, 10th Ed., Merck; Rahway, NJ

Final Draft: December 1994

Wright, W.H., and J.M. Schaffer, 1932. Critical Anthelmintic Tests of Chlorinated Alkyl Hydrocarbons and a Correlation Between the Anthelmintic Efficacy, Chemical Structure and Physical Properties. *Am. J. Hyg.* 16(2):325-428

Yurteri, C., D.F. Ryan, J.J. Callow, and J.J. Gurol 1987. The Effect of Chemical Composition of Water on Henry's law Constant, J. Water Pollut. Control Fed. 59, 950–956

Zoeteman, B.C.J., K.M. Harmsen, and J.B.H.J. Linders 1980. Persistent Organic Pollutants in River Water and Groundwater of the Netherlands, Chemosphere 9:231–249

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